



4everyone Functionality Kit 1000l

USER GUIDE

Check of real time PCR instrument functions

For research in vitro use only

SKU #2400-1001 4everyone Functionality Kit 100ul Includes low profile 100 µl PCR tubes, clear

The 4everyone Functionality Kit contains reagents and PCR tubes for 16 wells of a thermoblock to test the function of a real time PCR thermocycler.

Kit includes buffer and master mix which are necessary for running a real time PCR together with PCR tubes. The PCR tubes are containing specific primers and probes for a 2-channel real time PCR together with target DNA in different concentrations, all aliquoted in a dried format.

4everyone Functionality Kits are extremely temperature stable and ship at room temperature. Storage at 2-8 °C upon arrival is recommended.

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4everyone Functionality

This test checks the measurement capabilities of any 2-channel real time PCR thermoycler in a 200 μ L or 100 μ l tube format.

The 4everyone Functionality Kit is optimized but not limited to two-channel real time PCR thermocyclers using FAM channel (495/520 nm) and VIC/HEX channel (530/550 nm) to verify the correct function of both the hardware and software of the thermocycler.

Whenever there are unspecific results from a PCR test and it is unknown if the failure originates from a hardware error or from false sample handling, you can check the correct function of your thermocycler instrument by just running a functionality test. It is especially useful after moving the instrument or after a calibration was done. With the Functionality Kit, you can easily verify that your sample DNA is measured correct and the software read-out gives out the results within the correct concentration ranges, too.

Section 2

Introduction to the 4everyone Detection Kit Technology

Today, the use of PCR is accepted as the standard method for detecting nucleic acids from numerous microorganisms in a diversity of food and beverages, both functional species as well as spoilers. Real time PCR is one of the most powerful, specific and reliable methods for the quantitative detection and identification of microorganisms at an early process stage to prevent spoilage and to maintain overall product quality.

The 4everyone Detection Kit system is based on DNA amplification and detection of microorganisms by real time PCR. The specific PCR reagents, primers and probes, come in a ready-made dried format in the PCR tubes for unrivalled ease of use and temperature stability.

All PCR tests use the FAM channel (495/520 nm) for detection of the target microorganisms and the VIC/HEX channel (530/550 nm) for an internal control reaction. This allows 4everyone Detection Kits to prevent false negatives due to sample inhibition, allowing you to be truly confident about negative results.

A typical workflow includes the following four steps:



Kit Components

The 4everyone Functionality Kit contains sufficient reagents for 16 reactions.

Kit material for DNA isolation and analysis	Amount	Storage
Rehydration buffer B (blue cap)	1 x 275 μL	
2 x Master mix (white cap with blue sealing ring)	1 x 264 μL	dark at
PCR tubes (strips of 8) with mix of primers and probes	2	2-8 °C
Cap strips (strips of 8) for covering the PCR reaction tubes	2	

Table 1: Material supplied

Section 4

Shelf Life and Storage

Once received, the kit must be stored at 2-8 °C in the dark. Unless storage and/or time in transit at conditions above 8 °C wasn't exceeding 14 days in total, all reagents can be used until the best before date as indicated on the package label.

Section 5

Material Required but Not Supplied

Equipment	Supplies
Real time PCR thermocycler for 0.1 or 0.2 mL tubes with detection channels for FAM (520 nm emission) and VIC/HEX (550 nm emission)	Pipette tips with filters
Centrifuge for 8-tube strips 0.1 or 0.2 mL or adaptor for benchtop microcentrifuge	Gloves, powder free
Reaction tube mixer (Vortexer) (optional)	
Microliter pipettes	
100-1,000 μL variable volume	

Table 2: Additional material required

Detailed Instructions

Warning! Read the manual and the Safety Data Sheets before starting the analysis. Safety Data Sheets are available in the download area from our website https://pika-weihenstephan.com. All handling steps should be performed under sterile conditions. Wear appropriate protective clothing and powder free gloves. The use of filter tips is recommended.

DNA Analysis by real time PCR

Each of the PCR tubes in a Functionality Kit contains all components for a two channel PCR reaction in a dried format, including the target and internal control DNAs. The only exception is master mix which needs to be added as a reconstitution solution.

The target DNA is contained in different amounts per tube which will result in detection of different concentrations, while the internal control is inculded in equal amounts in all PCR tubes. PCR will give negative and positive results in channel 1 (FAM), showing three different concentrations in the positive results. All tubes should give equal results for the internal control reaction which is measured in channel 2 (VIC/HEX). The results are read out automatically using the automated read-out of results from the Open qPCR software.

The expected results are shown in the following table:

	A1	A2	A3	A4	A5	A6	A ₇	A8	
Α	(high)	valid negative (NTC)	medium	low	low	medium	negative (NTC)	high	Н
	B1	B ₂	В3	В4	B5	В6	В7	В8	
Α	high	negative	medium	low	low	medium	negative	high	Н

Table 3: Results per tube from 4everyone Functionality Kit

Preparation and Distribution of the Reconstitution Solution for PCR

- 1. Prepare the reconstitution solution by pipetting 264 μ L of Rehydration buffer B (blue cap) into the delivered vial 2 x Master Mix (white cap with blue sealing ring)
- 2. After adding the Rehydration buffer, re-close the reaction tube with the white cap which now contains the reconstitution solution. Mix briefly by vortexing or inverting the tube a couple times. Follow up with a quick spin down to collect the liquid at the bottom of the tube
- 3. Pipet 30 µL of the reconstitution solution into each of the 16 PCR tubes
- 4. Close all PCR tubes with the provided cap strips
- 5. Spin down the PCR tubes shortly (10-15 seconds) to collect the liquid at the bottom (max. 2,000 rpm), and check for trapped air bubbles
- 6. If trapped bubbles are present, repeat step 5.
- 7. Transfer all PCR tubes into the thermocycler and follow instructions according to the instrument's software

Instrument and Software Setup

Set up a PCR test with the following temterature and time protocol.

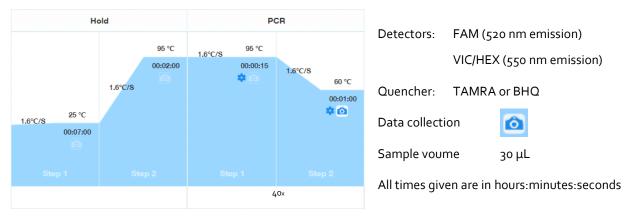


Fig. 1: Temperature scheme of thermocycler

Data Analysis

- 1. Follow user manual of thermocycler instrument
- 2. Evaluate the thermocycler results
 - Verify the curves
 - Expected values for the different included DNA concentrations Ct values are given in table 4
 - FAM channel detects two üarallels of all target DNAs in 3 different concentrations plus the negative control
 - VIC/HEX channel detects internal positive control reaction in all PCR tubes, expected value is Ct 30-33 for all tubes, independent from FAM value

Concentration	high	medium	low	negative
Expected Ct value in FAM channel	10.00-23.99	24.00-29.99	30.00-38.00	> 38.00

Table 4: Expected FAM channel results for different PCR tubes - manual evaluation

Use of Chai Open qPCR thermocycler

- 1. Select "RUN A TEST KIT"
- 2. Select Kit manufacturer "PIKA"
- Select "Single spoiler detection"
- 4. Select "Omit Positive Control"
- 5. Assign wells for experiment with different DNA concentrations as shown below
- 6. Run test



Fig. 2: Assigned wells for Open qPCR experiment with omitted Positive Control

Data Analysis with Open qPCR

All data can be analyzed directly at the end of the PCR run or at a later time by opening the stored experiment. Follow instructions of the corresponding Open qPCR software manual for opening experiments and setting the data analysis parameters.

Once the run is completed, the software displays the full data interpretion in a table format, an example is given in table 5. Full data including amplification curves can be seen in 'Full Results' view

Single spoiler detection

Well	Sample		Result	Cq	Quantity	
A2	Negative Control	\oslash	Valid			\Box
A 3	medium	\oplus	Positive	28.97	Medium	\Box
A4	low	\oplus	Positive	31.92	Low	\Box
A 5	low	\oplus	Positive	31.79	Low	\Box
A6	medium	\oplus	Positive	29.17	Medium	\Box
A7	negative	\ominus	Negative		Not Detectable	\Box
A 8	high	\oplus	Positive	19.04	High	\Box
B1	high	\oplus	Positive	19.03	High	\Box
B2	negative	\ominus	Negative		Not Detectable	\Box
В3	medium	\oplus	Positive	28.95	Medium	\Box
B4	low	\oplus	Positive	32.18	Low	\Box
В5	low	\oplus	Positive	32.00	Low	\Box
B6	medium	\oplus	Positive	29.24	Medium	\Box
В7	negative	\ominus	Negative		Not Detectable	\Box
В8	high	\oplus	Positive	19.09	High	\Box

Table 5: Results from Chai Dual Channel Open qPCR™ thermocycler

Precautions and Recommendations for Best Results

- ✓ This test must be performed by trained persons
- ✓ All potentially infectious material should be autoclaved before disposal
- ✓ The quality of results depends on strict compliance with Good Laboratory Practices (for example, the EN ISO 7218 standard), especially concerning PCR:
 - The laboratory equipment (pipets, tubes, etc.) must not circulate from one workstation to another
 - It is essential to use positive and negative controls for each series of amplification reactions
 - o Do not use reagents after their expiration date
 - Periodically verify the accuracy and precision of pipets and the correct functioning of the instruments
- ✓ Change gloves often, especially if you suspect they are contaminated
 - Clean work spaces periodically with at least 5% bleach or other DNA decontaminating agents such as DNA AWAY
 - Use powder-free gloves and avoid fingerprints and writing on tube caps as this can interfere with data acquisition

Section 8 Appendix

Trademarks and Property Rights

Trademarks:

FastOrange and PIKA Weihenstephan are registered trademarks or trademarks of PIKA Weihenstephan, Pfaffenhofen, Germany, in Germany and other countries.

Use of product:

4everyone Detection Kit is to be used for in vitro research purposes only.

Property Rights:

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"No license under these patents to use the PCR process is conveyed expressively or by implication to the purchaser by the purchase of this product. Further information on purchasing licenses to practice the PCR process may be obtained by contacting the Director of Licensing at Roche Molecular Systems, Inc., 1145 Atlantic Avenue, Alameda, California 94501."





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